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DEPARTMENT OF MINES SOUTH AUSTRALIA

GEOLOGICAL SURVEY PAEYNCLOGY SECTION

S.A.G. COOTANOORINA NO. 1 WELL
UPPER PALAEOZOIC AND LOWER CRETACEOUS MICROPALAEONTOLOGY

Grid G/3

bу

W. K. Harris Palynologist

and

B. McGowran Assistant Senior Palaeontologist

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W.K. HARRIS PALYNOLOGIST

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ASSISTANT B. McGOWRAN

ASSISTANT SENIOR PALAMONTOLOGIST

PALAMONTOLOGY SECTION

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ABSTRACT

Microfossils (spores, pollen, microplankton, foraminifera) from the section intersected in Cootanoorina No. 1 Well have been analysed biostratigraphically and environmentally.

Age determinations include: Unit 3 (2925 - 3111ft.), Devonian; Units 2 and 1 (1695-2925ft.), Lower Permian; Mount Toondina Beds (615-1595ft.), Lower Permian; Algebuckina Sandstone (467-615t.), no fossil evidence; Cadna-Owie Formation (370 - 467ft.), Lower Cretaceous (Valanginian to Aptian); Bulldog Shale (0 - 370ft.), Lower Cretaceous (Valanginian to Aptian).

The presence of foraminifera and organic-walled microplankton indicates marine influence during the deposition of Unit 1 in the Lower Permian. Below 2200ft. in Unit 2, the foraminiferal fauna is diverse though entirely arenacecus, and microplankton occur consistently. It is suggested that deposition occurred in a barred basin with rising temperatures and salinity. Anhydritic sediment above 2200ft. coincides with the sudden disappearance of foraminifera. Fossil evidence indicates that during deposition of the blackshale facies above 1745ft. bottom conditions were stagnant and saline, with a surface layer of fresher water. The Mount Toondina Beds are a paludal, fluviatile and lacustrine complex; no marine fossils were found.

INTRODUCTION

This report summarizes an investigation of microfossils from the sequence encountered in Cootanoorina No. 1 Well, drilled by the South Australian Department of Mines in the Arckaringa Sub-Basin (Latitude 28°00'30" South, Longitude 135°20' East).

Text-figure 1, showing the position of the well in

relation to other subsurface sections and the approximate limits of Permian subcrop, is taken from Wopfner (1964, fig. 1). It is seen that Cootanoorina No. 1 is further north, and further "offshore", than any other section, with the exception of the (stratigraphically restricted) section at Mt. Toondina (Freytag 1965). Thus, it could be expected that study of the section would be quite rewarding; and this has been justified. Wopfner and Allchurch (1967) have already discussed the significance of this well relative to the petroleum potential of the Arckaringa Sub-Basin.

Most attention was devoted to the Palaeozoic and particularly the Permian part of the sequence, and the fossil groups considered are pollen, spores, organic walled microplankton and foraminifera.

The aims of the study were correlation and age determination of the various rock units and an analysis of the Permian depositional environments.

Upper Palaeozoic biostratigraphy in South Australia is still in its infancy. That Devonian is here proven for the first time, is an indication of this. Since Permian in this region was first recognized on fossil evidence a decade ago (Balme 1957) there have been some advances palynologically, but a major attack on the many problems has still to be made. Foraminiferal studies have been taken further by Ludbrook (1967b), but study of the local restricted faunas is hampered by a lack of detailed comparative criteria from regions of greater faunal diversity.

Santos Oodnadatta No. 1 Well, about 50 miles northeast of Cootanoorina (Fig. 1), has been used as primary reference for the Mesozoic section. Dettman (1963) has studied the microfloras and Ludbrook (1966) the foraminifera from the Oodnadatta section. In the case of microfloras and microplankton, further comparisons are made with the type sections

of Mesozoic formations and also the sections drilled near Mt. Toondina.

W.K. Harris carried out the palynological and B. McGawran the foraminiferal investigations.

METHODS

Although rotary cuttings had to be used extensively, particularly for the foraminiferal studies, there are some indications that contamination has not significantly distorted the recovered assemblages. Depths given are uncorrected; corrections are insignificant. Bit samples were taken at 1580, 1736 and 1902ft. for palynological examination. For the details of drilling and the lithological sequence reference is made to the Well Completion Report by Allchurch and Wopfner (1967, unpub.).

Foraminifera and megaspores were recovered by standard methods of sample disintegration and washing. Selected megaspores were bleached with a mixture of sodium hypochlorite and concentrated by hydrochloric acid followed by a short treatment with 10% ammonium hydroxide. Smaller pollens and spores were recovered by a method similar to that described by Balme and Hassell (1962).

Representative specimens of the larger microfossils, foraminifera and megaspores, were photographed in reflected light using a Leitz Utropak Lighting unit and objectives mounted on a Leitz Laborlux microscope. Bleached megaspores, smaller palynomorphs and thin sections of foraminifera were photographed with conventional objectives on the Laborlux. Photomicrographs were recorded on 35mm. Adox KB14 film.

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kindly granted permission to refer to his unpublished studies.

Miss R. Phelan carried out the final stages in producing the

plates.

DEVONIAN

Unit 3 (2925-3111ft.)

A microflora frem a grey clay shale at 3010ft. contains Geminospora lemurata, Gramulatisporites cf. G.phillipsi, Convolutispora fromensis, Emphanisporites sp., Retusotriletes sp., Reticulatisporites textilis, ?Diaphanospora sp. and ?Leiozonotriletes.

No other fossils were recovered from this interval.

de Jersey (1966) has most recently summarised and
compared sustralian Devonian microfloras. Assemblages of this
age have been described from the Canning Basin (Balme and
Hassell 1962), the Carnarvon Basin (Balme 1960), the Amadeus
Basin (Hodgson 1964 unpub.) and the Adavale Basin (de Jersey
1966).

whilst the Cootanoorina assemblage is small and poorly preserved, it does have affinities with the described Devonian microfloras. In particular, it closely resembles those from the Gneudna correlatives in the Carnarvon Basin and the Parke Siltstone in the Amadeus Basin. For the present the age of the unit can be regarded as middle to upper Devonian.

PERMIAN

The following units, distinguished on their lithological and geophysical characteristics (Allchurch and Wopfner, 1967 unpublished; Wopfner and Allchurch 1967), have been dated biostratigraphically as Permian:

Mount Toondina Beds (615-1695ft.)

Unit 1 (1695-2550ft.)
Unit 2 (2550-1925ft.)

DISTRIBUTION OF FOSSILS

Feraminifera

The distribution of Permian foraminifera is summarised in text - fig. 1 and more fully in table 1.

Foraminifera were not found below 2750ft. Occasional specimens of six species were found in cuttings samples between 2600 and 2750ft. It is possible that they represent caving from 2300-2400ft., but this is thought unlikely even though none were found in core 6 at 2697ft. Three cuttings samples between 2420 and 2600ft. yielded no fauna.

Foraminifera occur consistently in cuttings between 2420 and 2200ft. However, two samples from core 5 between 2356 and 2359ft. appear to be barren. The specimen count drops from a maximum at 2200ft. to zero at 2190ft.

Occasional, poorly preserved specimens of

Lagenammina and Thuramminoides were found in two samples from
core 4, between 1745 and 1750ft., and in cuttings from 1730
and 1980ft. No fauna was found in the Mt. Toondina Beds which
were examined at 50ft. intervals.

Note on Foraminiferal Systematics: The systematics of Permian foraminifera from South Australia recently have been studied by Ludbrook (1967b). Identification of the Cootanoorina faunas is made primarily with reference to this work; comparative material from the Australian Permian as a whole (Crespin 1958) has not been available.

Preservation of the Cootanoorina faunas is rather poor, due to flattening, breakage and strongly adherent sapropelic matrix, especially above 2300ft. Consequently, identification is often tentative and sometimes impossible. The discrimination of species and genera at present is based largely on gross form of the test (shape, direction of growth)

with wall structure subordinate. When wall structure is analysed more thoroughly, using numerous thin sections, the information content of distorted and fragmented specimens is expected to increase.

Examples of probable confusion include: Glomospirella nyei with Hemidiscus balmei; Hyperammina hebdenensis with H. acicula and H. coleyi; Digitina recurvata with Textularia bookeri and Recurvoides wilsoni. The latter example is important because Digitina recurvata is much more common than in other Permian assemblages from this State (see below); furthermore, its morphological aspect at Cootanoorina (pl. figs. 15-20) differs from previous records (Crespin 1958, Ludbrook 1967b).

The identifications on text-fig. 2 and table 1 should be considered with these points in mind. It is felt, however, that the charts are quite accurate as to species numbers and general faunal trends.

Characteristics of Foraminiferal Fauna: Relative abundances (table 1) are based on an exhaustive search of approximately the same amount of each washed cuttings sample. Counts of specimens and species between 2450ft. and 2190ft. are graphed, on this quasi-statistical basis, in text-fig. 3. The specimen trend would be accentuated if significant contamination has occurred. That it has not, is suggested by a distinct change in preservation between 2280ft. and 2300ft. This change matches the lithological change from uniform dark grey, silty sapropelic shale downwards to an interbedding of this sediment type with light grey, sandy, plastic shale.

The richer samples have been plotted percentagewise for the four higher taxa distinguished here (text-fig. 3) following Loeblich and Tappan (1964). The superfamily Lituolacea includes the multichambered genera Digitina, Textularia, Recurvoides, Reophax and Ammobaculites.

The family Ammodiscidae includes Ammodiscus,

Hemidiscus, Tolypammina, Glomospira, Ammovertella and Ammovertellina. The family Astrorhizidae includes Hippocrepinella and Hyperammina. Use of these family-group taxa avoids two taxonomic problems. One concerns the identification of species, as discussed above. The other is that certain genera, particularly among the Ammodiscids, are based on rather doubtful criteria.

The important characteristics of the fauna are:

- 1. Its entirely arenaceous (agglutinated) character.
- 2. An increase upward in species and specimen numbers to 2300ft., with continued increase in specimens but decrease in species to 2200ft.
- 3. General decrease in Astrorhizidae numbers and increase in Lituolacea, the Ammodiscidae and Saccamminidae changing less. However, Ammodiscus sonahensis makes up 30% of the assemblage at 2200ft.
- 4. The Lituolacea are dominated by <u>Digitina recurvata</u> which makes up 58% of the total assemblage at 2250ft. and 52% at 2200ft.
- 5. Other variations in distribution and abundance (table 1) are relatively minor. There is no convenient criterion for recognizing more than one successional assemblage.

Microplankton

Organic walled microplankton referable to the Acritarcha (Downie et.al 1963, Segroves, 1967) were recovered from most samples in both Units 1 and 2. Species of Leiosphaera occur most consistently and abundantly in some samples together with Quadrisporites horridus. The upper limit of Q. horridus is in core 4 at 1,743ft. and the lowest occurrence is at 2,770ft. The most notable feature about its occurrence is that it is only associated in this well with proven planktonic genera such as Leiosphaera, and foraminifera, suggesting strongly that the species is a marine planktonic organism, perhaps

tolerant of marginal marine conditions. Other acritarchs noted but of rare and restricted occurrence include <u>Leiofusa</u> sp. in core 5 at 2,359ft. and poorly preserved <u>Michystridium</u> sp. in core 5 at 2,697ft.

Megaspores

Single specimens, probably from cavings, are recorded from 1,980-1,990ft. and 2,090-2,100ft. and will not be considered further. The first significant occurrence is at 1,920ft. - 1,930ft. where the assemblage consists of twenty specimens of the genera Carruthersiella and Triletes. Throughout the remainder of the Permian sequence to 630ft.

Carruthersiella dominates the assemblages. There is possibly some significance in the first appearance of Mammilaespora at 950ft. At higher levels in the Mount Toondina Beds the genus is co-dominant with Carruthersiella. The only lageniculate trilete specimen was recovered from core 3 at 1,243ft. and is referred to the genus Dijkstraea. The genus Duosporites does not appear to show any marked changes in distribution but it might be noticed that it does not occur in assemblages with Mammilaespora above 950ft.

sediments of the world, megas pores from Australian Permian sediments have received little attention. They have been recorded frequently in foraminiferal washings as "lycopod megaspores" by Ludbrook (1961, 1967a). Høeg, Bose and Manum (1955) recognised the importance of examining megaspores in transmitted light and elucidating internal structures such as the mesosporium, its ornamentation and its attachment to the exosporium. Following these authors, Pant and Srivastava (1961, 1962) examined Gondwana megaspores paying particular attention to the number of walls, their attachment and ornament. The classification and nomenclature of Pant and Srivastava is adopted in this report. It must be realised however, that many more specimens will have to be bleached and examined before any

finality on generic subdivisions can be achieved. No attempt has yet been made to relate the Cootanoorina forms specifically with described species. However, it can be noted that the majority of forms do differ from Permian species described from India, Africa and South America.

Miospores

The lowest occurrence of Permian miospores occurs in cuttings at 2,830ft. Parasaccites spp., Cycadopites cymbatus, Protohaploxypinus geraiensis and occasional trilete spores including Punctatisporites gretensis, and Vallatisporites sp. characteristise the assemblage. This assemblage changes little through Unit 2 but adding progressively more components of the striate bisaccate genera. Cores 6, 7 and 8 within Unit 2 failed to yield any miospores. The assemblages are, characteristically, not diverse, and there is little ather organic debris in the residues apart from occasional microplankton.

within Unit 1 the assemblages become more diversified in respect to the bisaccate and trilete spore component.

Parasacites spp. remain prominent and towards the top of the unit G. cymbatus becomes very common. Apiculatisporis levis,

Verrucosisporites pseudoreticulatus and Ovalipollis sp. first make their appearance; V. pseudoreticulatus at 2,100-2,110ft.

The distribution of miospores across the Mt. Toondina

Beds - Unit 1 boundary show no significant qualitative changes.

In character the microfloras of the lower sections of the

Mt. Toondina Beds becomes dominated even more by striate

bisaccate pollen.

In core 3, Striatoabietites fusus, Vesicaspora maxima and Marsupipollenites triradiata make their first appearance, the latter species occurring rarely. In contrast to other Permian sequences in the Lake Phillipson Region Marsupipollenites spp. do not reach a high proportion in the microfloral assemblages so far examined. Camptotriletes biornatus and Krauselisporites apiculatus appear in coal

cuttings at 830ft. and continue to the top of the sequence.

Core 2, the highest Permian sample examined, yielded a particularly diverse and well preserved assemblage dominated by bisaccate pollen (especially the larger forms; Platysaccus leschiki and Vesicaspora maxima), apiculate and acanthine trilete species, and cingulate mesosporoids. Parasaccites spp. still remain prominent in the assemblage. Of some importance is the occurrence at 633ft. of a species referable to Bascanisporites undosus.

Other Pranic Remains

Specimens of several species of Permian foraminifera contain sponge spicules in the test agglutinate. The spicules are mostly pyritised.

The anhydritic sequence between 1900ft. and 2200ft., apparently barren of foraminifera, contains "abjects" which appear to be organic in origin although thin sections show them to be structureless internally. They may be worm casts.

ENVIRONMENT

The analysis of Permian environments and their change in this profile through the Arckaringa Sub-basin can be divided conveniently into (a) Units 2 and 1, which are discussed in some detail, and (b) Mount Toondina Beds.

Units 2 and 1: Devonian-Permian

Contact to base Mount Toondina Beds

The environmental analysis of fossil assemblages includes both the actualistic and the comparative, biofacies approaches. The first must be used with some caution here because of the great evolutionary changes of extinction and relay in foraminiferal faunas since the Palaeozoic. Mesozoic to Recent foraminiferal assemblages tend to become more arenaceous in their content as a response to any or all of the following environmental stresses: higher or lower salinity, lowered temperature, lowered oxygen, increased turbidity,

acidity. This tendency will be less evident in the Palaeozoic where smaller species with secreted-calcareous tests never reached the degree of dominance in normal shelf assemblages later achieved by the lamellar foraminifera. Thus Permian assemblages tend to be more arenaceous than their younger counterparts. A comparative approach accordingly is heightened in value. Detailed studies of faunal distributions, their changes, and their relation to other groups, particularly shelly megafossils, are needed for the Gondwana sequences.

Clues to the environment in which the Cootanoorina sequence from 2925ft. to 2200ft. was deposited include: first appearance of foraminifera and microplankton at 2700-2750ft.; a general but sporadic increase in foraminiferal specimen and species numbers; the arenaceous but rather diverse nature of this fauna; the absence of a calcareous, shelly fauna; lithological evidence for a change from glacial to evaporitic conditions.

Features of the Unit 2 sequence (graded bedding, intraformational slumping, dolomitic beds, resistivity characters together with undisputed glacial sediments along the eastern margin of the Boorthanna Trough have led Wopfner and Allchurch (1967) to envisage remobilisation of glacigenes by density currents down a submarine slope. A marine environment (sensu lato) is supported by the presence of both foraminifera and microplankton. Foraminifera can live in very cold waters; their apparent absence below 2750ft. is explained, rather, by the coarse nature of the clastics, particularly since no specimens were found in core 6 or core 5 both of which contain microplankton.

On the other hand, the sporadic distribution of foraminifera between 2750ft. and 2400ft., where lithology is not an obvious control, followed by their consistent occurrence in some numbers and diversity up to 2300ft., indicates increasingly marine influence. That is, the sequence is transgressive,

although it may have been "marine" throughout.

Ludbrook (1956-1967b) has stressed the restricted, sparse nature of local Permian faunas. In so far as calcareous fossils are almost entirely absent, this is so. However, the foraminiferal fauna of about 20 species in the richest sample (2300ft; table 1) compares quite favourably with arenaceous assemblages described from the Upper Palaeozoic. Assemblages recovered by washing techniques tend to contain 20-30 species in the Mississippian and Pennsylvanian of North America, occasionally with more and often with less (for example: Plummer 1945, Conkin 1961, Conkin, Conkin and Pike 1965). Acid residues (for example, Ireland 1956) yield comparable numbers, with better recovery of delicate arenaceous specimens but destruction of calcareous tests. Crespin (1958) has studied assemblages from the Byro Group and Noonkanbah Formation of the Carnarvon and Canning Basins respectively. In her lists of "characteristics" species in largely or entirely arenaceous assemblages there are always less than 20 species. It is not clear whether these lists refer to the total species numbers found. Against this, the lists refer to entire formations, so that numbers are broadly comparable with the Cootanoorina assemblage. Many of these Australian and North American assemblages are associated with rich, shelly, marine megafaunas.

marine, if not open marine, is also indicated by comparison with Recent and fossil assemblages from marginal marine environments (estuary, lagoon, marsh). Although these assemblages are predominantly arenaceous, they are characterised by low diversity and by the dominance of very few species (Phleger 1960, Walton 1964) Fossil examples include: the Ammobaculites—

Haplophragmoides fauna in a regressive part of the Midway Group in Texas (Kellough 1965); the coastal facies in the Cretaceous of northern Alaska (Tappan 1960) with only one or two species per sample but locally abundant specimens; marine ingressions in the Westphalian of Germany (Bartenstein 1950)

with monotonous arenaceous assemblages; a <u>Glomospira</u> - <u>Hyperammina</u> assemblage in the Permian of Tasmania, high in the Ferntree Group and preceding the non-marine Cygnet coal Measures (Conkin and Conkin 1965, unpublished).

When considered in the light of these examples of marine and marginal marine faunas, the Cootanoorina fauna at its level of maximum diversity (2300ft.) appears to indicate a fully marine environment. Hence, the problem is to explain a severely restricted but internally rather diverse fauna. Assemblages overlying glacigenes are known from the Quamby Mudstone in Tasmania and the Holmwood Shale in the Perth Basin, W.Aust. (Crespin 1958). In both cases the species are all arenaceous and diversity is rather low (about 10 species among several samples). In the similarly placed Grant Formation (Fitzroy Basin) Crespin has only five. The situation is similar in the Permian of South Australia (Ludbrook 1956-1967b). There is some evidence that the number of arenaceous species in Recent shelf assemblages increases at very high latitudes (Saidova 1966). Thus the Cootanoorina assemblage can be explained as a marine fauna colonizing a cold environment.

Obviously, however, this explanation does not cover the continued absence of calcareous foraminifera and shelly megafossil remains while the temperature rose to an evaporitic level. Another restriction must have operated. Physical restriction of the basin leading to evaporitic conditions (see below) appears to be foreshadowed in the composition of the fauna. Other evidence for this is found in the faunal change in the dark, sapropelic, silty shales between 2300 and 2200ft. In this interval specimen numbers tend to increase and species numbers to decrease so that Ammodiscus conshensis and Digitina recurvata together account for more than 80% of the specimens at 2200ft. (text-fig. 3). The small numbers involved warrant caution, but these trends together would suggest regression (Walton 1964). Either or both raised salinity and lowered

oxygen supply (but not anaerobic conditions) can account for the trends. It is noteworthy that "euxinic" conditions are suggested for the deposition of the Bulgadoo Shale and Quinnanie Shale in the Byro Group, Carnarvon Basin (Thomas 1958, McWhae et al. 1958). Foraminiferal lists for each formation appear to show no significant drop in diversity below normal marine assemblages (Crespin 1958). However, detailed studies of sections probably would yield valuable comparative information since the Quinnanie Shale contains a single-species assemblage of Hyperammina coleyi.

The sudden and complete disappearance of faraminifera but not microplanktan above 2200ft. is consistent with the onset of evaporitic conditions.

The siltstones above 2080 feet no lorger contain anhydrite; they are replaced above 1945ft. by dark, carbonaceous shales. The disappearance of microplankton at about 1700ft. overlaps the lowest accurrence of megaspares at about 1900ft.; foraminifera are very rare and sporadic in their occurrence above 2000ft. Resistivity characters indicate broadly marine salinities. All this evidence combines to suggest the advance of a paludal environmental complex toward the depositional area. Increased runoff terminated anhydritic deposition, brought in megaspores, and eventually drove out microplankton. It also formed a surface layer of fresher water, thus lowering circulation with bottom waters retaining their high salinity. Occasional foraminifera indicate very slight marine influence; possibly they were transported.

In conclusion, the sequence represented by Unit 2 and Unit 1 can be interpreted as due to a complex of changing factors operating in and around a trough, which may have been barred throughout the time span involved. Ludbrook (1967b) has suggested the existence of fjords in this part of the Great Artesian Basin. The present evidence does not contradict this, but the origin by faulting of the Boorthanna Trough makes it

unlikely. The Boorthanna Trough and Arckaringa Sub-Basin formed a gulf which became for a time normal marine, in a sense, but not open marine.

Local Comparisons: In terms of diversity, specimen abundance and preservation the best foraminiferal faunas known so far from South Australia are in the Engineering and Water Supply Department Stuart Range No. 3 subsurface section (text-fig. 1).

A comparison of Cootanoorina No. 1 with Stuart Range No. 3 is summarised in text-fig. 3. The comparison is restricted to intervals in which foraminifers are relatively common: Cootanoorina, 2200-2350ft., 7 samples; Stuart Range, 1182-1367ft., 9 samples. Data for the latter are based on samples assembled and reported by Ludbrook (1967a) Permian foraminifera are sufficiently rare to justify the assumption that no personal bias is involved in recovering either assemblage.

There are no significant differences between the two sections in presence-or-absence of species. The main differences in relative abundance are: (1) generally more Hyperamminids at Cootanoorina. (2) More Saccamminids at Stuart Range; the group is dominated by Saccammina orca. (3) Ammodiscus oonahensis is more common at Stuart Range. (4) The Lituolacea are dominated in the higher part of the Cootanoorina section by Digitina recurvata. This species is relatively rare at Stuart Range where Recurvoides relatively rare at Cootanoorina, is one of the most important species.

Below 2280ft. at Cootanoorina, mixed preservation indicates that foraminifera occur in both the dark shales and the interbedded pale grey, plastic shales. Specimens from the latter are similar to the Stuart Range specimens in preservation. Also <u>Digitina recurvata</u> is more similar morphologically to specimens from Stuart Range than those at 2280ft. and above. Finally <u>Recurvoides vilsoni</u>: occurs mostly in the lower part of the Cootanoorina section.

It would seem, then, that there are two biofacies at

Cootanoorina, distinguished largely by the abundance and form of <u>Digitina recurvata</u> in the dark, silty shales. The dark shale assemblage is the more divergent of the two from the Stuart Range assemblage and also from most of the other fossiliferous Permian in this State, as judged from inspection of all samples assembled by Ludbrook (1956-1967). The only exception occurs at Boorthanna (text-fig. 1; see Ludbrook 1961). Here, assemblages are sparser and more poorly preserved than at Cootanoorina, but there are indications that the change between 1800 and 2088 feet is similar. <u>Digitina</u> of the Cootanoorina type occurs at 1800ft; <u>Recurvoides</u> of the Stuart Range type at 2088ft; the preservation is of dark, sapropelic shale type at 1800-1950ft. but mixed at 2000-2088ft.

These comparisons indicate that faunal differences are due to environmental changes rather than differences in age. The dark shale facies at Boorthanna and Cootanoorina, underlying the anhydritic sediments in the latter section is due to formation of a barred basin in this region, whereas barring is not manifest in sequences described from the Lake Phillipson trough (Ludbrook 1961). Further studies of foraminiferal distribution as more sections become available should make these biofacies differences more meaningful.

Mount Toondina Beds

Allchurch and Wopfner (1967, p.25) concluded on the evidence of abundant plant fossil material, numerous coal beds, the high resistivity and the general lithological character of the sediments that the formation was of continental origin; paludal, lacustrine and fluviatile environments were recognised. The abundance of megaspores and the absence of marine fossils support this conclusion.

CORRELATION AND AGE

General Discussion: The biostratigraphic standard for subsurface sequences such as the present one is based on changes in microfloral assemblages. Accordingly, these are treated in most detail here, with contributory evidence from the foraminifera.

With the exception of a recent paper by Segroves (1967) no systematic studies of Australian Permian microfloras have been published since the series of pioneering papers by Balme and Hennelly (1955, 1956a, 1956b). In these papers some indications were given as to the usefulness of Permian microspores in correlation. Subsequently Balme (1964) produced a paper on the Australian pre-Tertiary floras in which he named the "Striatites-Microflora" as being characteristic and perhaps synonymous with the Permian. This microflora he divided into three assemblages, two in the lower Permian, the Nuskoisporites-Assemblage and the Vittatina-Assemblage, and the Dulhuntyispora-Assemblage in the upper Permian. Whilst this is the first formal naming of the units, they were already in use, as evidenced by Balme's paper on the Lake Phillipson Bore (1957). With regard to relating these units to the accepted Permian divisions, Balme relied on correlations with marine invertebrate assemblages, some containing ammonoids, from sequences in the Canning, Carnarvon and Perth Basins. Thus the Nuskoisporites-Assemblage is dated as Sakmarian and Lower Artinskian, the Vittatina-Assemblage as Artinskian and the Dulhuntyispora-Assemblage as Upper Permian.

Evans (1967 unpub.) has further resolved the Nuskoisporites and Vittatina Assemblages into two divisions each, using an informal nomenclature but giving them Stage status. Thus, Evans uses Stages 1 (the oldest) to 4, with his Stage 5 being equivalent to Balme's <u>Dulhuntyispora</u> Assemblage. The specific identity of many of the key fossils in the stages is, however, not clear and a systematic section in the paper would have been most useful. As a consequence there is some difficulty in recognising the Stages precisely. Although Evans discusses the limits of the Permian system on the basis of

plant macrofossils and consequent microfloral expressions, he avoids any discussion of terms such as Sakmarian and Artinskian.

The correlation of Permian microfloras from Cootanoorina are discussed in relation to these two papers. It is however noted that finer divisions are possible in a more local setting using cores from the Lake Phillipson bore as a standard. Megasperes would appear to have some stratigraphic utility on a local scale. The results of this study as yet not completed, cannot be discussed in this report.

Unit 2: Miospores from this unit are referred to Stage 2 of Evans characterised by <u>Parasaccites</u> spp. and rare striate bisaccate species, <u>Punctatisporites gretensis</u> and <u>Vallatisporites</u> sp. and the unit is therefore of lower Permian age. Balme (1963, p. 59) states that the lower part of the glacial sequence in Western Australia is not rich in plant microfossils. This is true also for similar sequences in the western portion of the Great Artesian Basin.

Assemblages from this unit resemble closely those from the Lake Phillipson bore at about 2300ft. Although Evans (1967) regards these as representing Stage 1, examination of samples at this level has yielded bisaccate pollen of the <u>P. goraiensis</u> type and an assemblage of Stage 2. Unit 2 is therefore a correlative in part of the Lake Phillipson Beds (Ludbrook 1967b).

Unit 1: There are no perceptible qualitative changes in the microfloral component across the Unit 1-Unit 2 boundary. The absolute number of misspores does increase however as does the proportion of organic detritus. The appearance of spores characteristic of Stage 3 occurs at about 2100ft. In particular V. pseudoreticulatus makes its initial appearance at this level.

Stage 3 according to Evans, corresponds to the lower portion of the <u>Vittatina Assemblage</u>. The prominent genus of the <u>Vittatina Assemblage</u>, <u>Marsupipollenites</u>, does not reach significant frequencies in the assemblages and it is here suggested that Stage 3 may be in part correlated with the <u>Nuskoisporites</u>

Assemblage of Sakmarian age. The age of Unit 1 is therefore regarded as Sakmarian.

Unit 1 is correlated in part with the Stuart Range Beds (Ludbrook 1967b) and it is of some correlative value to note the appearance of megaspores within this unit throughout the region. Table 2 summarises the distribution of megaspores and their relationship to foraminifera in some bores in the area.

Table 2

Bore Name	Megaspores	<u>Foraminifera</u>
Cootanoorina No. 1	650-1900ft.	1730-2750ft.
Boorthanna Bore No. 1	185- 400ft.	1800-2088ft.
Stuart Range No. 3	507- 815ft.	740-1620ft.
Stuart Range No. 1	485- 668ft.	-

It is clear that there is some overlap between megaspores and foraminifera and in this region megaspores appear at a particular time later in Unit 1 or in the Stuart Range Beds. Ludbrook's (1967a) correlation of portion of the Stuart Range sequence with the Mt. Toondina Beds is not substantiated palaeontologically or lithologically. In particular two specimens of Thuramminoides sphaeroidalis (one at 740-745ft. and one at 750-755ft.) and the associated megaspores indicate a correlation with the upper portion of Unit 1.

Mt. Toondina Beds: Stage 3 assemblages continue into the base of this formation and the first appearance of Camptotriletes biornatus at 830ft. together with Verrucosisporites parmatus and Apiculatisporis cornutus marks the onset of Stage 4 of Artinskian age. The highest assemblage from Core 2 is unlike any from the lake Phillipson Bore and the presence of PBascanisporites undosus, very common large bisaccate pollen and echinate species similar to and including Didecitriletes ericianus suggest that Stage 5 is present. The absence of Dulhuntyispora and Gnetaceaepollenties sinuosus places some restriction on this interpretation. Evans (1967 p.8) suggests

that Stage 5 commences in the Lower Permian.

The age of the Mt. Toondina Beds is therefore Artinskian with some possible extension into the Sakmarian.

Correlation of the well section of the formation and the outcropping type section is based on one sample only. Although more than twenty samples were collected and processed, only one, a dark brown carbonaceous clay, yielded recognisable miospores. The sample is from Freytag's Unit 16 (1965, Fig. 4) and yielded the following assemblage:

Acanthotriletes cf. 1. filiformis A. tereteangularis Apiculatisporis levis Granulatisporites micronodosus Krauselisporites apiculatus Laevigatosporites vulgaris Marsupipollenites triradiata Microfoveolatispora directa Neoraistrickia ramosa Ovalipollis sp. Punctatisporites gretensis Parasaccites spp. Platysaccus leschiki Retusotriletes diversiformis Schizo poris scissus Striatiti undiff. Vesicaspora ovata

V. maxima

Verrucosisporites pseudoreticulata

The assemblage shows distinct similarity to Stage 3 microfloras occurring below 830ft. in Cootanoorina No. 1. As there is some distance ($4\frac{1}{2}$ miles) between the type section and the well a precise correlation in terms of coal seams is not attempted. It is apparent that the outcrop section is more eroded at its top in comparison with the well section.

Foraminiferal Evidence: Strata containing Permian microfaumas in South Australia have been dated as Lower Permian, Sakmarian, and Lower Sakmarian (Ludbrook, 1956-1967b). The clearest evidence for correlation, however, is palynological as discussed above.

The foraminiferal fauna of Yorke Peninsula was compared (Ludbrook 1956, 1965) with that from the Quamby Group in Tasmania, of Sakmarian age, but also (Ludbrook 1956) with that from the Mulbring Siltstone of the Maitland Group in New South Wales, of Upper Permian Age. Foraminifera have been used to make rather broad correlations within the Australian Permian (see particularly Crespin, 1958); and Conkin and Conkin (1965, unpublished) recently have presented preliminary results leading toward a zonation of the marine Permian in Tasmania. In both cases calcareous (secreted) foraminifera are available, thus adding significantly to the biostratigraphic potential of the group. Within South Australia no more than one assemblage, or "zone", can be recognised in known species ranges. Indeed, an assemblage of twelve species from the Eucla Basin was dated as Artinskian on palynological evidence, yet does not differ significantly from Sakmarian faunas found elsewhere in the State (Harris and Ludbrook 1966).

The Cootanoorina fauna is of Lower Permian aspect rather than Upper Permian. The presence of Ammodiscus oonahensis is evidence for a Sakmarian instead of Artinskian age. So far as is known (Crespin 1958, Conkin and Conkin 1965, unpublished) this species is limited to the Sakmarian.

Ludbrook (1967b) records A. conahensis from the Eucla Basin fauna, but the specimens are considered here to be distinct and conspecific with Ammodiscus of multicinctus as recorded herein.

MESOZOIC

Algebuckina Sandstone (467-615ft.)

Cuttings from this unit in the well were considered unsuitable for palynological examination. No foraminifera were found in three cuttings samples. The unit occurs in the Oodnadatta well below 1100', and Dettman regards this interval as falling within the Speciosus ssemblage and therefore of Lower Cretaceous age. This determination can be accepted realising that it is based solely on rotary cuttings (see Ludbrook 1966, p. 27). It is significant that the nominate species of the Stylosus Assemblage was not recorded in the well. It is acknowledged that the unit may be in part Jurassic in the deeper parts of the Basin (Wopfner, 1964). At present there is no fossil evidence for this supposition.

Cadna-Owie Formation (370-467ft.)

No foraminifera or other fossils were found in the two cuttings samples washed and examined.

Cuttings from this unit were not processed for palynology, but it is worth recording here a microfloral assemblage recovered from the type section of the formation, 2.5 miles southwest of Algebuckina Railway Siding, Algebuckina 1-mile sheet.

A very dark grey carbonaceous clay (Unit 8) 38ft.

from the top of the Cadna-Owie Formation yielded a well
preserved and diverse microflora. The assemblage is as follows:

Aequitriradites spinulosus

Alisporites grandis

A. similis

Ceratosporites equalis

Coronatispora perforata

Cicatricosisporites australiensis

Classopollis sp.

Cyathidites australia

C. asper

Cyclosporites hughesi

Contignisporites cooksonii

<u>Dictyotosporites</u> speciosus

Foraminisporis wonthaggiensis

Gleicheniidites cf. G. circinidites

<u>Ischyosporites punctatus</u>

Leptolepidites verrucatus

Lycopodiumsporites austroclavatidites

L. nodosus

Microcachryidites antarcticus

Osmundacidites wellmanii

Podocarpidites ellipticus

Rouseisporites reticulatus

The assemblage is dominated by an abundance of gymnosperm pollen, in particular \underline{A} . similis and \underline{M} . antarcticus.

Correlation and age: The presence of both <u>C. hughesi</u> and <u>D. speciosus</u> identifies the assemblage with the lower portion of the Speciosus Assemblage of Dettman (1963). Correlatives of this unit occur between 743 and 1292ft. in Santos Oodnadatta Bore No. 1 and between 1330 and 1472ft. in Cootabarlow Bore. On the basis of the apparent limited vertical distribution of <u>Coronatispora perforata</u>, which has not been recorded from any of the Bulldog Shale samples (see below), a finer subdivision gives the following correlation:-

Cootabarlow Bore between 1354 and 1402ft.

Oodnadatta Bore No. 1 between 1502 and 1292ft.

Dettmann (1963) considers the age of the assemblage to be
Valanginian to intian.

In contrast to the younger Cretaceous units, the sample from the Cadna-Owie Formation did not yield any microplankton.

Microfloras and Microplankton: Microfloral assemblages have been recovered from Core 1 at 362ft. and 370ft. and from the well velocity survey shot holes S.F.4 at 85ft. and S.P.5, 88ft. The shot hole samples ("Bottom of hole samples") were examined to provide an upper limit to the age of the Bulldog Shale in the drilled section. Their proximity to Cootanoorina No. 1 permits reliable extrapolation to the well and assemblages are therefore recorded in the general palaeontological log of this well (fig. 1).

Assemblages were well preserved and diverse with a high proportion of organic walled microplankton. There are few specific differences between the sampled intervals.

Species of pollens and spores characteristic of the assemblage include Dictyotosporites speciosus, Cicatricosisporites australiensis, Cyclosporites hughesi, Contignisporites cooksonii, Trilobosporites perverulentus, Murospora florida (very rare), Lycopodiumsporites circolumenus and Pilosisporites notensis. Organic walled microplankton are represented by Canningia colliveri, Dingodinium cerviculum, Muderongia mcwhaei, Broomearamosa, Palaeotetradinium hyalodermum and Gonyaulacacysta edwardsi.

Foraminifera: No foraminifera were found in weathered material from 17ft. At 50ft., a poor arenaceous fauna is dominated by <u>Bigenerina loeblichae</u>.

From 100 to 300ft. the assemblages are mixed arenaceous and calcareous. The more common species include Bigenerina loeblichae, Trochammina minuta, Ammobaculoides romaensis, Textularia wilgunyaensis, Lenticulina spp. including forms referable to warregoensis Crespin and dalhousiensis Ludbrook, Praebulimina hergottensis, Anomalinoides mawsoni, and a species of Gavelinella which appears closest to G. parvula among the species described so far. All of these species, however, fluctuate considerably in their occurrence.

The lowest part of the Bulldog Shale carries an almost entirely arenaceous fauna. The main species are Textularia anacooraensis, Pseudobolivina engeninensis, Miliammina inferior, Trochammina raggatti. Again, however, there are variations in occurrence and abundance.

Correlation and Age: Ludbrook (1966) has erected four foraminiferal zones for the Lower Cretaceous stratigraphic sequence in the western part of the Great Artesian Basin. From top to bottom they are:

Neobulimina australiana

<u>Verneuilina howchini - Trochammina flosculus</u> <u>Hergottella jonesi</u>

Trochammina raggatti - Textularia anacooraensis

These "zones" are major biofacies units reflecting environmental fluctuation over a large region. Thus, the first (lowest) and third zones are based on arenaceous assemblages with the others containing a mixed calcareous-arenaceous fauna. At the same time each of the assemblages has its characteristic species so that these units, while probably allochronous, can be expected to become valuable in basin analysis.

The <u>Trochammina raggatti</u> - <u>Textularia anacooraensis</u> zone is recognisable at Cootanoorina below 350ft. As well as both name species, the species listed above are characteristic members of the assemblage.

The occurrence of <u>Hergottella jonesi</u> at 100ft. and 160ft. proves the presence of the <u>H. jonesi</u> zone, <u>H. jonesi</u> being restricted to its lower part (Ludbrook 1966). This is consistent with the relative abundance of calcareous species down to 300ft. and the presence of <u>Praebulimina hergottensis</u> and <u>Anomalinoides mawsoni</u>.

In Santos Oodnadatta No. 1 Well, <u>Verneuilina</u>
<u>howchini</u> occurs down to 570ft. and <u>Textularia anacooraensis</u>
first appears at 880ft. (Ludbrook 1966). Although the name

fossil was not found, the <u>Hergottella jonesi</u> zonal assemblage accordingly should occur between 570 and 880ft.; this is supported by the specific composition and calcareous nature of the faunas from this interval. A more precise direct correlation with the Oodnadatta section is not possible on this evidence, but if restriction of <u>H. jonesi</u> to the lower part of the zone is assumed, and if we further assume relatively little time-transgression of these biostratigraphic units between Cootanoorina and Oodnadatta, then a correlation of the Bulldog Shale at Cootanoorina with the "Marree Formation" below about 700ft. at Oodnadatta is indicated. The faunas from the Cootanoorina section support other evidence for the lack of Albian Oodnadatta Formation in this section and in the Mt. Toondina region generally (Freytag et.al. 1967).

On a regional basis there are two microfloral, blostratigraphic schemes in use for the Lower Cretaceous, those of Dettmann (1963) and Svans (1966). A comparison between the two is summarised in table 3.

TABLE 3

P.R. EVANS (1966)			M.E. DETTHINN (1963)
Characteristic Species	Unit	Unit	Characteristic Species
Coptospora paradoxa Trilobosporites trioreticulatus	K2b	Paradoxa Assemblage	C. paradoxa, Pilosisporites grandis Cicatricosisporites pseudotripartitus Trilobosporites trioreticulatus, T. tribotrys
Overlap of <u>C. paradoxa</u> and <u>Dictyotosporites</u> speciosus	K2a		Contigisporites glebulentus, Krauselisporites majus.
First appearance of <u>Crybelosporites</u> striatus with <u>D</u> . speciosus	K1 đ	Assemblage	D. speciosus, C. striatus, Pilosisporites notensis, Coptospora striata, Rouseisporites radiatus, Arcellites reticulatus
Cicatricosisporites australiensis, D. speciosus	K1 b-c	"Upper" 	D. speciosus, M. florida, Cyclosporites hughesi Contignisporites cooksonii, Krauselisporites
C. <u>nustraliensis</u> Cyclosporites hughesi D. <u>speciosus</u>		"Lower"	Contignisporites cooksonii, Krauselisporites linearis, Cooksonites variabilis.
Top of range of <u>Murospora</u> <u>florida</u>	K1 &	Stylosus Assemblage	Crybelosporites stylosus, Coronatispora perforata, Murospora florida, Biretisporites spectabilis, C. cooksonii.

clearly, the assemblage from the Bulldog Shale is equivalent to Dettmann's Speciosus assemblage, more precisely the lower portion, and is of Valanginian to Aptian in age. The assemblage is represented in Santos Oodnadatta No. 1 Well between 742 and 1292ft. Very similar assemblages were reported (Harris 1964 unpub.) from the Toondina No. 1 Stratigraphic Well which also penetrated a section of the Bulldog Shale. Hence the Bulldog Shale is of Valanginian to Aptian in age on microfloral evidence.

The microplankton component is dominated by

D. cerviculum and G. edwardsi. Evans (1966ap. 61) regards

these species as being mutually exclusive and thus providing

two microplankton divisions: "the lower marked by Dingodinium

cerviculum and Maderongia tetracantha and the upper by

Gonyaulax edwardsi and Diconodinium spp." He notes that the

D. cerviculum zone corresponds to the spore pollen units K1b-c,

and G. edwardsi occurs near the top of Unit K2 in the Otway

Basin (see Table 2). Clearly the ranges and factes relation—

ships of Australian Cretaceous microplankton need further study.

Alberti (1961 p. 6) compares G. orthoceras Eisenack with

G. edwardsi and notes a gradation between the two species, which

suggests that they may be synonymous. G. orthoceras ranges in

Europe from Barremian to Albian.

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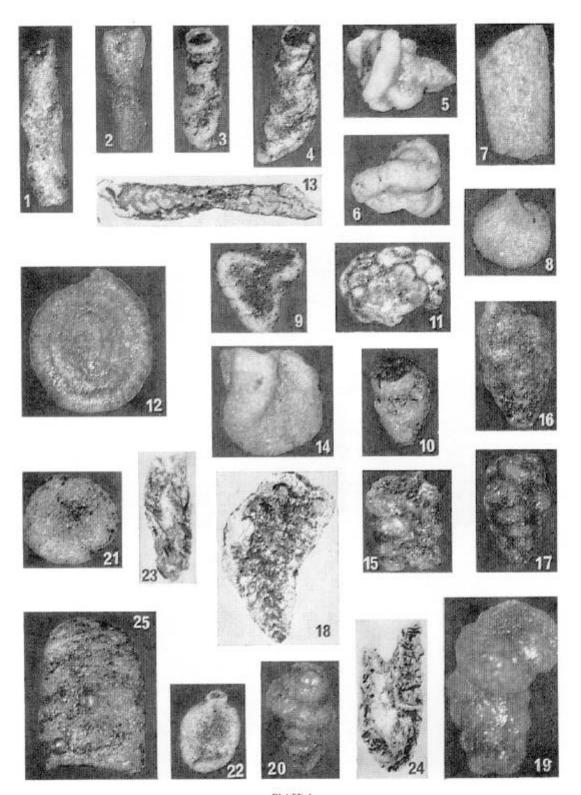


PLATE 1 83

PLATE 1

PERMIAN FORAMINIFERA

- Figs. 13, 18, 23-24, oriented thin sections in non-polarised transmitted light, Ca. x 50; other specimens, Ca. x 30 approx. All specimens from cuttings.
 - 1 2 Hyperammina acicula 1, fragment; 2, with initial end; both from 2230ft.
 - 3 4 <u>Hippocrepinella Diaperta</u>, showing characteristic wrinkling of test; both from 2250ft.
 - 5 6 Tolypammina undulata Both from 2300ft.
 - 7 8 <u>Hyperammina Lebdenensis</u> Two fragments, one an initial end; both from 2300ft.
- 9 10 Hyperammina expansa Showing variation in flare of test, although both distorted; both from 2250ft.
- 11 cf. Ammovertella Lowchini 2300ft.
- 12 13 Ammodiscus Oonalensis
 - 12, somewhat flattened; 2300ft. 13, showing sheartype distortion closing chamber, also tough, adherent, sapropelic matrix; 2250ft.
- 14 cf. Hemidiscus balmei 2300ft.
- 15 20 <u>Digitina Recurvata</u> and Parr. 15, 16, 18, 20, 2200ft.; 17, 19, 2300ft. Lack of clarity due to low contrast in sapropelic shale preservavation. 15, distal, biserial fragment, with large, incorporated quartz grains out of focus. 16-17, 20, showing <u>Eggerella</u> like triserial growth. 18, axial section showing poor condition of most tests; triserial (or initially multiserial) up to change in direction of growth, subsequently biserial.
 - 19, Large specimen, proximal end missing, with two large, biserially arranged final chambers; 2300ft.

- 21 Thuramminoides sphaeroidalis 2500ft.
- 22 Pelosina ampulla Flattened; 2250ft.
- 23 24 "Axial" sections of probable Saccammina area, showing incorporated, variously oriented, pyritized sponge spicules; both from 2300ft.
 - 25 Saccammina area. Flattened fragment; 2300ft.

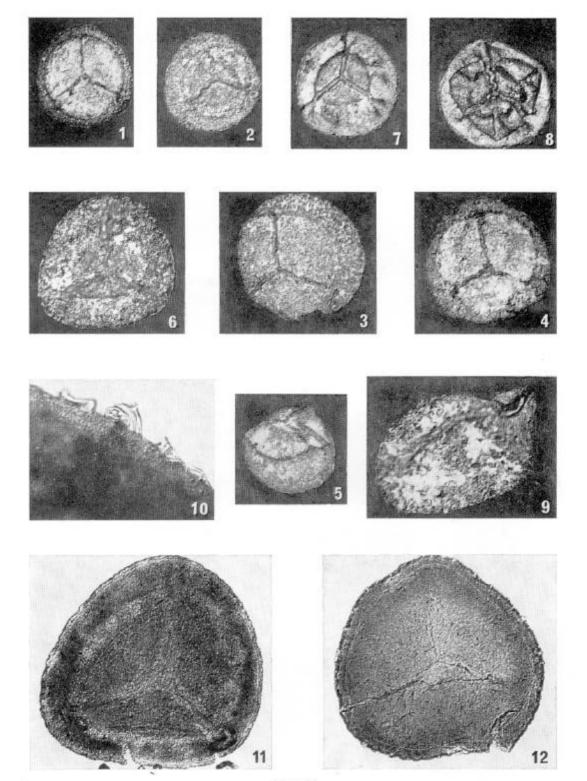


PLATE 2

PLATE 2 PERMIAN MEGASPORES

- Figs. 1-9, specimens photographed in reflected light Ca. x 50. Other figs. in transmitted light.
- 1 5, 11-12. Carruthersiella spp. Note the well contact areas, the wrinkled appearance of the aperture margins and the papillate exosporium. 1, 4, 5, 11812 in cuttings at 1000', 2 core at 1749'6", 3-cuttings at 950ft. 11, 12 Ca. x 125; note mesosporium in 11. Aperture margins and contact areas less distinct.
- 6, 10 <u>Manuilaespora</u> sp. Note, in 6, small contact area. 10, Ca x 500 note branded processes on the exosporium. Cuttings at 750ft.
- 7,8 <u>Duosperites</u> spp. Note prominent proximal attachment area of mesosperium and the nearly smooth surface of the exosperium. Core 3 at 1245ft., and core 4 at 1748 6" respectively.
 - 9. <u>Dighttraea</u> sp. Note lageniculate proximal surface. Distal surface ornamentation consists of dense branched processes similar to those in Mammilaespora. Core 3 at 1243ft.

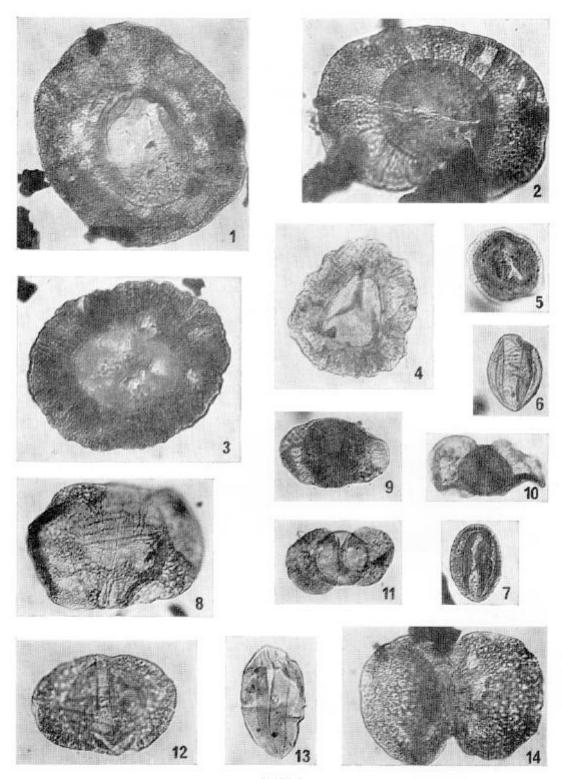


PLATE 3

PLATE 3

PERMIAN MIOSPORES

- Fig. 1, ca x 320, all other figs. ca. x 500.
- Plicatipollerites gondwanensis. Note trilete
 aperture and body infold system. Core 2 at
 633ft.
- ?Potoniesporites sp. Central body split. Core
 at 633ft.
- 3, 4. Parasaccites spp. Note in 4 the strongly upturned margins of the aperture and the sub-triangular shape of the central body. 3-from core 3 at 1244ft. and 4 from core 4 at 1743ft.
- 5. ?Bascanisporites undosus. Note reticulation of bladder. Core 2 at 633ft.
- 6, 7. Marsupipollenites triradiator. Core 2 at 633ft.

 Note striate form in 6 as against non-striate
 in 7.
- 8. <u>Protohaploxypinus gorgiensis</u> Outcrop sample,
 Mt. Toondina.
- 9. Ovalipollis sp. Note monolite scar on distal surface. Core 3 at 1244ft.
- 10, 11. Strictites cancellatus (Balme & Hennelly)

 Core 2 at 653ft. and Mt. Toondina outcrop sample respectively.
- 12. <u>Striates linupidus</u> (Balmo & Hennelly)

 Core 3 at 1244ft.
- 13. Cycadopites of G. cymbatus

 Core 2 at 633ft.
- 14. <u>Striatopodocarpidites fusus</u>

 Core 3 at 1244ft.

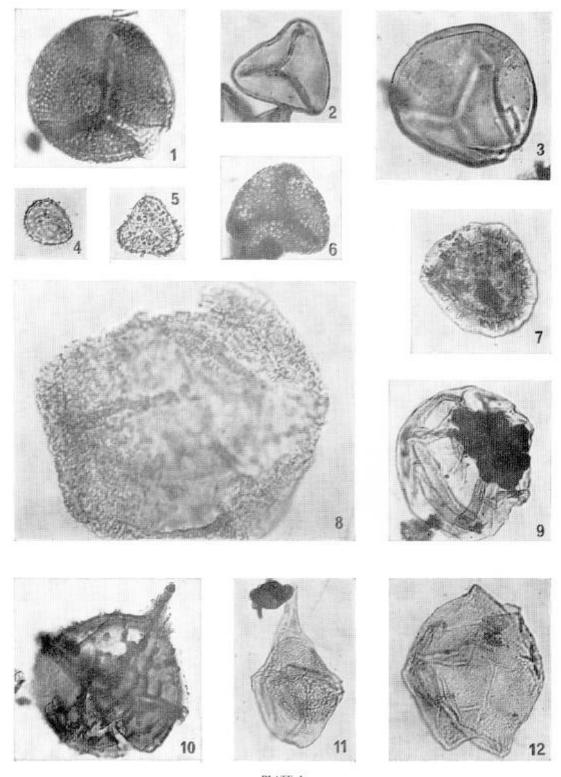


PLATE 4 89

PLACE 4

PERMIAN & DEVONIAN TRILETE MIROSPORES: PERMIAN & CRETACEOUS MICROPLANKTON.

All figures except fig. 8 ca. x 500. Fig. 8, ca x 1000

- 1. <u>Verrucosisporites</u> of <u>V. trisecatus</u>
 Outcrop sample, Mt. Toondina
- 2. <u>Microfoveolatispora directa</u>

 Core 2 at 653ft.
- 3. <u>Punctatisporites gretensis</u>

 Core 3 at 1244ft.
- 4. <u>Tuberculatosportites modicus</u>
 Outcrop sample, Mt. Toondina
- 5. <u>Neoraistrickia ramosa</u>

 Core 3 at 1244ft.
- 6. <u>Verrucosisporites</u> sp. Core 3 at 1244ft.
- 7. Krauselisporites apiculatus
 Cuttings at 850ft.
- 8. Geninospora lemurata

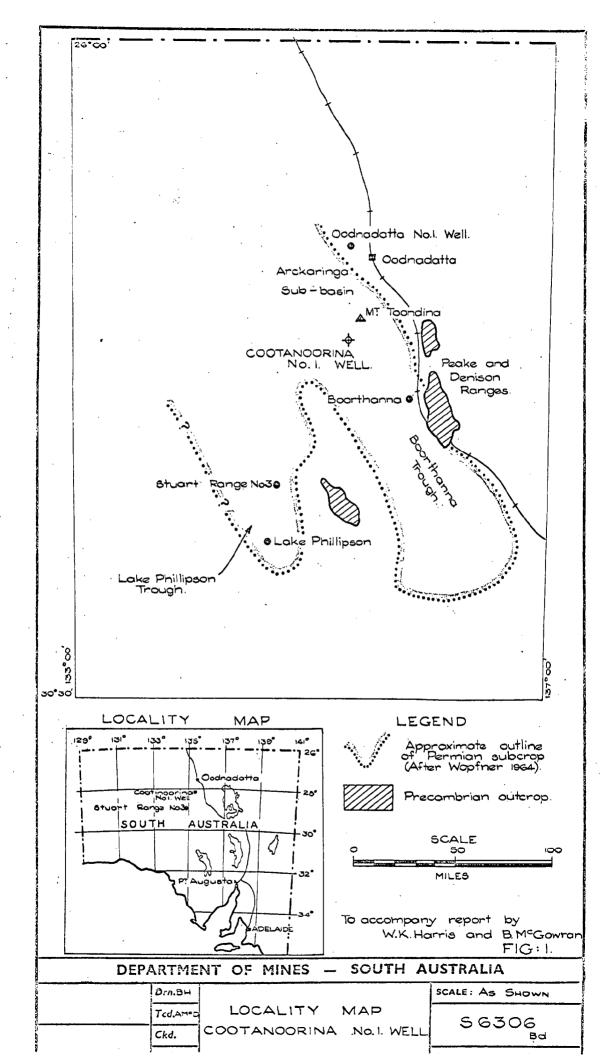
 Note mesosporium. Proximal face and ornament
 is also similar to <u>Retusotriletes</u> of <u>R. devonicus</u>

 Naumora cuttings at 3010ft.
- 9. <u>Leicsphaera</u> sp.

 Cors 6 at 2697ft.
- 10. <u>Genvanlandewsta edwardsi</u>

 Core 1 at 362ft.
- 11. <u>Dingodinium cerviculum</u>

 Core 1 at 3621t.
- 12. <u>Canningia colliveri</u>
 Core 1 at 362ft.



		CRETACEOUS	FORAMINIFERA	PERMIAN	FORAMINIFERA	MICROPLANKTO	N CRETACEOUS	MIOSPORES	MEGASPOR	RES P	ERMIAN - D	EVONIAN	MIOSPO	RES	_
Composite (Adapted from Wop	NTOLOGICAL LOG AND Well Log. Inter and Allchurch 1967) TIVITY TIVITY TIVITY TOUR AND LITHOLOGIC DESCRIPTION	igenerina loeblichae Crespin. Iaplophragmaides perturbans Ludbrook. Iaplophragmaides chapmani Crespin. Immobaculoides romaensis Crespin. Inmobaculoides romaensis Crespin. Inchammina minuta Crespin. Inchammina alalhousiensis Ludbrook. Iarginulinapsis animensis Ludbrook. Iarginulinapsis animensis Ludbrook. Iavelinella ef. parvula (Crespin). Inomalinaides mawson (Crespin). Inomalinaides mawson (Crespin). Inomalinaides mawson (Crespin).	Extularia wilgunyaersis Crespin. Seudonodosaria sp. Iorginulinopsis australis Crespin. Iorginulina marreensis Crespin. Iorginulina marreensis Crespin. Iorginulina marreensis Crespin. Iorginulina marreensis Crespin. Immobaculites australis (Howchin) etc. Seudobolivina engeninensis Ludbrook. enticulina warregoensis Crespin etc. extularia anacoaraensis Crespin Tochommina raggati Crespin Tiliammina infenior Ludbrook. Itiliammina infenior Ludbrook.	agenammina ampulla (Crespin). Incomminades sphaerodalis Plummer. Idomospira adhaerens Parr. Saccommina orca Ludbrook. Recurvoides wilsoni Ludbrook. Igitina recurvata Crespin and Parr. Extularia bookeri Crespin. Sularia bookeri Crespin.	demiciscus balmer Ludbrook. Impocrapinella bioperta Crespin. Impocrapinella bioperta Crespin. Impocrapinella bioperta Crespin. Impocrapinella bioperta Crespin. Impocrammina acicula (Parr.) Impocrammina expanso (Plummer). Imporia thomasi Crespin. Importa tricameratus Parr. Impocratellina glomospiroides Ludbrook. Immovertellina glomospiroides Ludbrook. Immovertellina glomospiroides Ludbrook.	eiotusa sp. eiosphaera sp. lichystridium sp. Juadrusporites horridus Hennelly. Palaeotetradinium hydlodermum Cookson. Dingodinium cerviculum Cookson and Eisenack. Sroomsa ramasa Cookson and Eisenack. Ponyaulacacysta edwardsi Cookson and Eisenack. Tuderangia mcwhaei Cookson and Eisenack. Syclosporites hughesi (Cookson and Dethmann)	Synthidites australis Couper. Stereisporites australis Couper. Stereisporites antiquasporites (Wilson and Webster.) Veoralistrickia truncatus (Cookson) Seratosporites equalis Cookson and Dethmann. Sycapodiumsporites australiensis (Cookson) Sicatricasisporites australiensis (Cookson) Sicatricasisporites australiensis (Cookson) Sicatricasisporites pudens Balme. Seticulatisporites pudens Balme. Seticulatisporites yearucatus Couper. Soraminisporis wonthaggiensis (Cookson and Dethmann.) Sictotosporites speciasus Cookson and Dethmann. Sictotosporites speciasus Cookson and Dethmann. Sullites tuberculiformis Cookson.	aevigatosporites ovatus Wilson and Webster. Sugaepollenites dampieri (Balme) T. trilobatus grandis (Cookson) T. trilobatus grandis (Cookson) T. Trilobatus anicrosaccatus (Cookson) T. Trilobatus ontarcticus (Cookson) T. Trilobatus ontarcticus (Cookson) T. Trilobatus australis (Cookson) T. Trilobatus australis (Balme) T. Trilobatus porites arcolumenus Cookson & Dettmann T. Tudbrooki Dettmann	Aquitriradites spinulosus (Cookson and Dethmann) Dilosisporites notensis Cookson and Dethmann. Tilobosporites perverulentus (Verbitskaya) Schizosporis reficulatus Cookson and Dethmann. Minerisporites sp. Mammillaespora sp. Triletes spp. Duosporites sp. Dijkstraea sp.	Pascanisporites undosus Balme and Hennelly. Cycadopites cymbatus (Balme and Hennelly). Aconthotriletes tereteangulatus (Balme and Hennelly). Injuditisporis levis Balme and Hennelly. Torinites eremus (Balme and Hennelly). Torinites eremus (Balme and Hennelly). Tranulatisparites micronodosus Balme and Hennelly.	Krouselisporites opicularius Jansonius. Marsupipollenites trirodota (Balme and Hennelly.) Micotoveolotispora directa (Balme and Hennelly.) M. pseudoreticulata (Balme and Hennelly.) M. trisina (Balme and Hennelly.) Veoraistrickia ramosa (Balme and Hennelly.) Porasoccites sp. Porasoccites sp. Plicatipollenites gondwanensis (Balme and Hennelly.)	Florysaccus rescriber mark. Potoniesporites sp. Purchatisporites gretensis Balme and Hennelly. C. Retusotriletes diversiformis (Balme and Hennelly) C. Striotoabietites multistriatus (Balme and Hennelly) Striotopoaccarpialites fusus (Balme and Hennelly) 5. phaleratus (Balme and Hennelly) Striatiti. undiff.	Tuberculariosporites modicus Balme and Hennelly Vallatisporites sp. Vallatisporites sp. Vallatisporites sp. Vallatisporites pormatus Balme and Hennelly. Vesicasporo moxima Hart. 1. ovata (Balme and Hennelly) 1. itatina scutata (Balme and Hennelly) 1. itatina scutata (Balme and Hennelly)	Cingulate mesosporoids undiff. Sonvolutispora framensis Balme and Hassell. Seminospora femurata Balme. Emphanisporites sp. ? Leiozonotriletes sp. Sramulatisporites cf. G. phillipsi de Jersey. Reticulatisporites textilis Balme and Hassell. Retusotriletes sp.	FORMATION
200	CALCAREOUS SANDSTONE offwhite to pale grey, shaley														CADNA- BULLDOG OWIE SHALE SET LOWER CRETACEOUS
800	SANDSTONE, pale grey coarse to very coarse quartz, some feldspar, subangular to sub-rounded, minor kaolinitic matrix. SILTSTONE, pale to dark grey micaceous, interbedded with CARBONACEOUS SHALE dark brown, and COAL, black, fibrous. Minor calcareous sandstone.									•••		• • • •	• • • •		NA BEDS ALGEBUCK SANDSTON TO TO TO TO THE PROPERTY OF THE PROP
140Q 1600	SANDSTONE, pale grey, very fine to fine grained. SILTSTONE, pale grey, clayey, slightly calcareous and carbonaceous. Some thin CALCAREOUS SANDSTONES.									• • •		• • • •			MOUNT TOONDI
2000 SCA	SHALE, dark grey, laminated, slightly silty and carbonaceous. SILTSTONE, mid-grey with specks of white serkite up to 0.5mm. dia. laminated. Anhydritic between 2080 and 2190 ft. ALE 30 NGE-75 SILTY SHALE, dark grey, micromicaceous, slightly calcareous.										• • • •	• • • •			UNIT I LOWER PERN
2600 2800 2800 2800	SANDY SHALE, pole grey, plastic when wet, slightly calcareous, interbedded with SHALE dark grey, non-plastic, and thin calcareous sandstones, offwhite, pebbly. SANDSTONE, pole grey fine grained pebbly to conglomerate in places, interbedded with SHALE, grey, plastic when wet, and thin dolomite, brown to black, crystalline, vuggy.									•	• • • · · · · · · · · · · · · · · · · ·	••••	•		S TINO
3000	DOLOMITE, pale grey sandy, interbedded with DOLOMITIC SHALE, green-grey and ANHYDRITE pink.									To ac	ccompany Pal. Re	eport 1/68 by	W. Harris and	B. Mc Gowran.	UNIT 3
											. J	<u>, </u>		ES — SOUTH AUS	STRALIA
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7 1.

COMPOSITE WELL LOG

Drn. V., H. SCALE: AS SHOWN

Tcd. A MGD

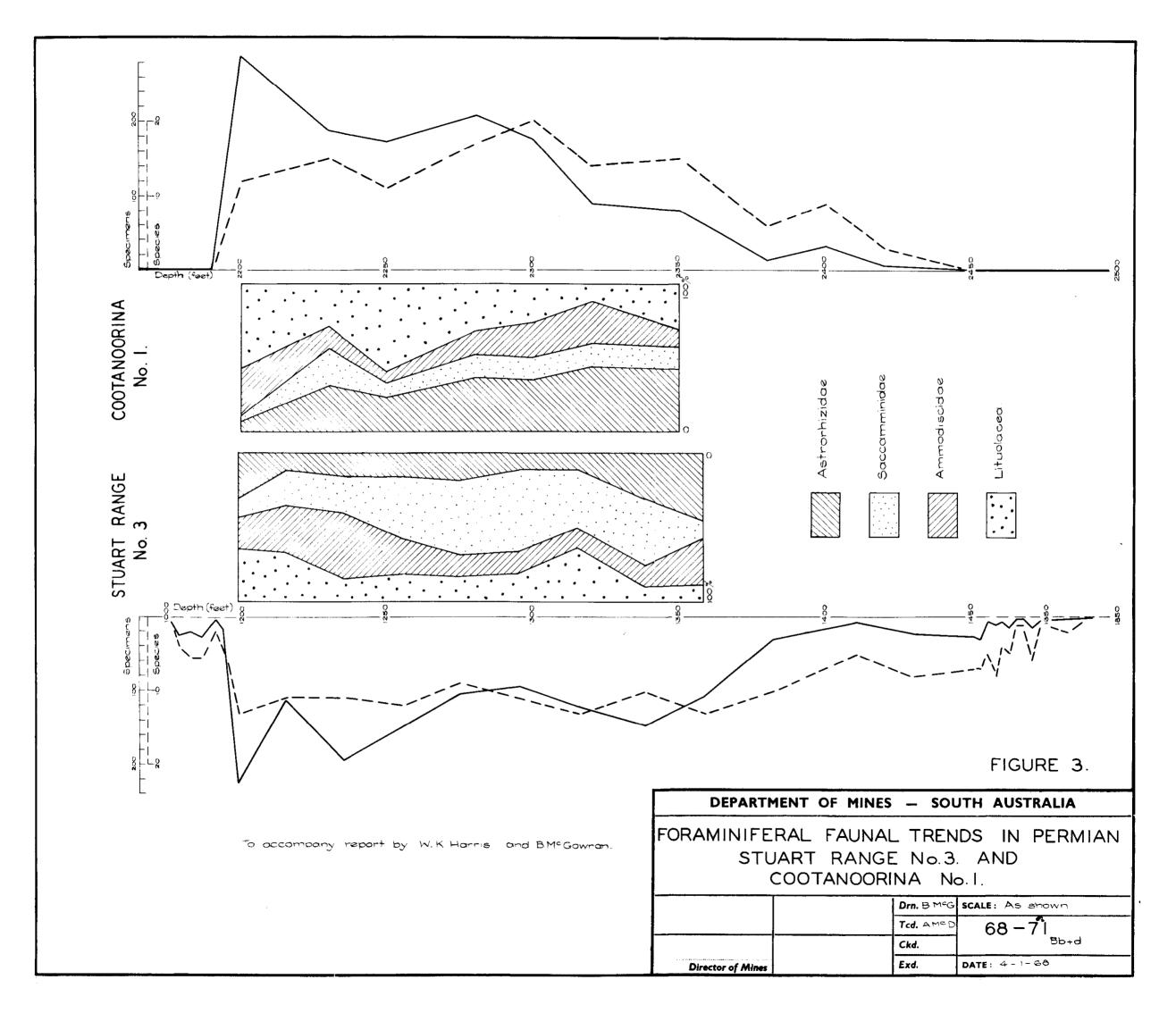
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Director of Mines Exd. DATE: 16-1-68



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